

Susceptibility of Clinical Isolates of *Staphylococcus aureus* to Ceftaroline

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ABSTRACT

Introduction: Methicillin-Resistant *Staphylococcus aureus* (MRSA) infection is a major global healthcare problem, the prevalence of which varies from 25-50% in India. It is known to cause Skin and Soft tissue Infections (SSI), endovascular infections, endocarditis, pneumonia, septic arthritis, osteomyelitis, and sepsis. Vancomycin is the drug of choice for treating severe MRSA infections. Ceftaroline, a fifth-generation cephalosporin has been approved by the United States Food and Drug Administration (US FDA) for treating acute bacterial SSI caused by susceptible micro-organisms including MRSA, Community acquired respiratory tract infection, MRSA bacteremia and endocarditis.

Aim: To assess the susceptibility of clinical isolates of *S. aureus* to ceftaroline, in a Tertiary Care Hospital.

Materials and Methods: This prospective study was conducted in the Department of Microbiology of a Tertiary

Care Hospital over a period of two months from June 2019 to July 2019. *S.aureus* isolates from various clinical samples were screened for methicillin resistance by disc diffusion method using cefoxitin disc and ceftaroline susceptibility of these isolates was assessed by E-strip method. The isolates were classified as ceftaroline susceptible, Susceptibility Dose Dependent (SDD) and ceftaroline resistant respectively as per CLSI guidelines. A descriptive analysis of the data was done and the results were presented as frequencies and percentages.

Results: All the *S.aureus* isolates were found to be susceptible to ceftaroline. Methicillin Sensitive *Staphylococcus aureus* (MSSA) isolates had lower Minimum Inhibitory Concentration (MIC) when compared to MRSA. The highest MIC among MRSA was 0.5 µg/mL.

Conclusion: Ceftaroline can be considered as an effective alternative for treatment of infections caused by MRSA.

Keywords: Ceftaroline susceptibility, Methicillin resistant *Staphylococcus aureus*, Minimum inhibitory concentration

INTRODUCTION

The MRSA infection is a major global healthcare problem, the prevalence of which varies from 25-50% in India [1]. It is known to cause Skin and Soft tissue Infection (SSI), endovascular infections, endocarditis, pneumonia, septic arthritis, osteomyelitis, and sepsis [2]. Vancomycin is the drug of choice for treating severe MRSA infections. However, the use of vancomycin has been associated with several limitations which include poor penetration of the drug into the tissues, narrow therapeutic index, slow bactericidal activity, difficulty in achieving pharmacokinetic/pharmacodynamic targets and potential side effects like nephrotoxicity and ototoxicity [3,4]. Also, a meta-analysis has reported treatment failures with vancomycin therapy in critically ill patients which may be attributed to suboptimal therapeutic levels or high MIC values [5]. Alternative drugs like linezolid, daptomycin are being used increasingly for treatment of MRSA infections [6].

Ceftaroline, a fifth-generation cephalosporin has been approved by the US FDA for treating acute bacterial SSI caused by susceptible micro-organisms including MRSA, community acquired respiratory tract infection, MRSA bacteremia and endocarditis [7]. This antimicrobial inhibits cell wall synthesis by binding to Penicillin Binding Proteins (PBP) 1, 2, 3 and PBP 2a for MRSA [8]. Clinical trials have shown that ceftaroline is well tolerated by patients [9]. Also, it has been shown to be as effective as vancomycin, daptomycin and linezolid in eradicating MRSA [9,10]. Resistance to ceftaroline is not very common. Several studies have reported decreased susceptibility of MRSA to ceftaroline in sporadic cases [11,12]. The resistance may be due to the mutation within PBP 2a protein, in particular, outside the Penicillin- Binding Domain (nPBD) [7].

In India, there are very few studies undertaken to evaluate the susceptibility of *S.aureus* to ceftaroline and there is a limited data about the susceptibility pattern of *S.aureus* to ceftaroline [13-15]. Therefore, this study was conducted to screen *S.aureus* isolates

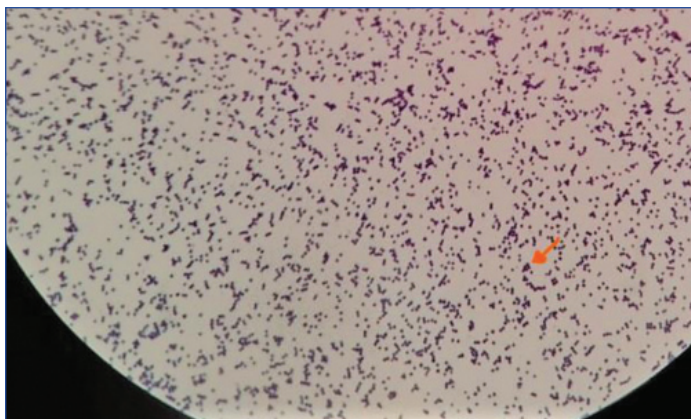
obtained from various clinical samples for methicillin resistance and assess their susceptibility to ceftaroline, in a Tertiary Care Hospital.

MATERIALS AND METHODS

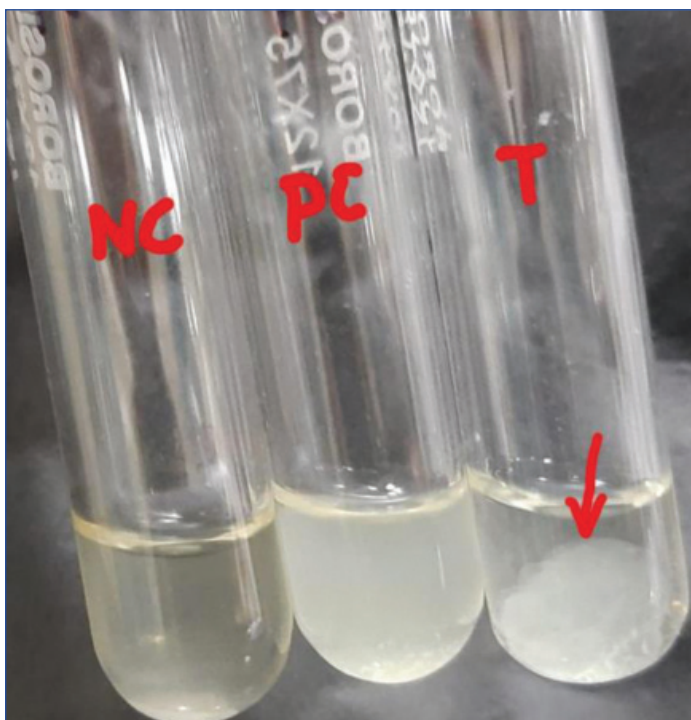
This prospective study was conducted in the Department of Microbiology over a period of two months from June 2019 to July 2019 as part of ICMR-STs 2019 (Reference ID.2019-02280) Clearance was obtained from the Institutional Ethics Committee (INST.EC/EC/057/2019-20).

Fifty non-duplicate *S. aureus* strains isolated from various clinical samples were included in the study. The strains were streaked on nutrient agar plates after thawing the vials. A smear was prepared from an isolated colony and stained with Gram's stain [Table/Fig-1]. Biochemical tests like catalase test and tube coagulase test were done to reconfirm the identity of the strain. Gram positive cocci which were catalase positive and tube coagulase positive [Table/Fig-2] were identified as *Staphylococcus aureus*.

Screening for MRSA: Screening for methicillin resistance was done by modified Kirby Bauer disc diffusion method using cefoxitin (30µg) discs [16]. Inoculum for lawn culture was prepared by direct colony suspension method. Three to five isolated colonies of *S. aureus* were suspended in 5 mL of peptone water and the turbidity of the test suspension was standardised to match 0.5 McFarland standard. A lawn culture of the test organism was made on Mueller Hinton agar plates according to standard protocols. Cefoxitin disc (30 µg) was placed on the lawn culture and the plates were incubated at 35°C for 16-18 hours. The diameter of the zone of inhibition was measured using a ruler. A zone size of ≥22 mm was interpreted as methicillin sensitive and ≤21 mm was interpreted as methicillin resistant as per Clinical and Laboratory Standards Institute (CLSI) guidelines. *S. aureus* American Type Culture Collection (ATCC) 25923 and *S. aureus* ATCC 43300 were used as controls [16].



[Table/Fig-1]: Gram positive cocci in singles, pairs and clusters. (magnification- X1000)



[Table/Fig-2]: Tube coagulase test. The arrow showing positive tube coagulase yielded by the test isolate. NC: Negative control, PC: Positive control; T: Test isolate

Ceftaroline susceptibility: Testing for ceftaroline susceptibility was done by E-strip method. The ceftaroline E strips 0.002-32 µg/mL was obtained from Biomerieux, France. Inoculum preparation and lawn culture of the test organism was done as detailed for disc diffusion method. The E-strips were placed on the lawn culture and the plates were incubated at 37°C for 18-24 hours. MIC's were read where the ellipse intersects the MIC scale. Since E-strip has continuous gradient, MIC values "in-between" two-fold dilutions can be obtained. These values were rounded up to next two-fold dilution before categorisation. MIC≤1 µg/mL, 2-4 µg/mL and ≥8 µg/mL were interpreted as ceftaroline susceptible, SDD and ceftaroline resistant respectively as per CLSI guidelines. *S. aureus* ATCC 29213 was used as control [16].

STATISTICAL ANALYSIS

Descriptive analysis of the distribution of sample, age, gender, and antimicrobial susceptibility data were done, and the results obtained were presented as frequencies and percentages.

RESULTS

Among the 50 *S.aureus* isolates 28 (56%) were isolated from male patients and 22 (44%) from females. The age range of patients from whom the *S.aureus* was isolated was 2-73 years, mean age being 41 years.

Of the 50 non-duplicate *S. aureus* isolates, 47 (94%) were isolated from pus, two (4%) from blood and one (2%) from endotracheal aspirate.

Among the 50 isolates tested 28 (56%) were methicillin resistant and 22 (44%) were methicillin sensitive. The age and gender distribution for MRSA and MSSA are shown in [Table/Fig-3].

Age group (in years)	MRSA		MSSA	
	No. of Males	No. of females	No of Males	No of females
0-20	3	0	2	1
21-40	3	8	3	2
41-60	5	4	8	4
61-80	2	3	2	0
Total	13	15	15	7

[Table/Fig-3]: Age and gender distribution for MRSA and MSSA.

All the *S. aureus* isolates were found to be susceptible to ceftaroline with their MIC's ranging from 0.064 to 0.5 µg/mL. Frequency distribution of ceftaroline MIC's of MSSA and MRSA isolates are shown in [Table/Fig-4]. It was also observed that the MSSA isolates had lower MIC's when compared to MRSA. Majority (90%) of MSSA isolated had an MIC of ≤0.25 µg/mL in comparison to 13 (46.43%) of MRSA isolates which had an MIC of ≤0.25 µg/mL. A MIC of 0.5 µg/mL was observed among 15 (53.57%) of the MRSA isolates which included 14 isolates having MIC of 0.38 µg/mL rounded-up to 0.5 µg/mL.

Ceftaroline MIC (µg/mL)	No. of MSSA isolates (%)	No. of MRSA isolates (%)
0.064	3 (13.64)	0
0.125	11 (50)	0
0.25	6 (27.27)	13 (46.43)
0.5	2 (9.09)	15 (53.57)
Total	22	28

[Table/Fig-4]: Frequency distribution of ceftaroline MIC's among MSSA and MRSA isolate.

A MRSA isolate with ceftaroline MIC of 0.25 µg/mL is shown in [Table /Fig-5].



[Table/Fig-5]: MRSA isolate with ceftaroline MIC of 0.25 µg/mL.

DISCUSSION

Staphylococcus aureus is an important cause of hospital as well as community acquired infections. In the era of increasing antimicrobial resistance, treating the infections caused by MRSA is posing a real challenge to the clinicians. Emergence of multi-drug resistant MRSA isolates further complicates the treatment of infections caused by these organisms [13]. Ceftaroline fosamil has been approved as an alternative for a severe MRSA infection [8].

In this study, all the 50 clinical isolates of *S. aureus*, inclusive of 22 MSSA and 28 MRSA strains were found to be susceptible to ceftaroline. This correlates with the findings of an Indian study, where all the 50 MRSA isolates isolated from various clinical samples were found to be sensitive to ceftaroline [13]. Similar results were also observed in a multi-centre study from Spain, where the all *S. aureus* isolates tested were inhibited by ceftaroline with a MIC of ≤ 1 $\mu\text{g/mL}$ [17]. The antimicrobial resistance surveillance program, Assessing Worldwide Antimicrobial Resistance and Evaluation (AWARE), which evaluated the trends in *S. aureus* susceptibility rates to ceftaroline, reported a 100% susceptibility to ceftaroline among MSSA. However, the study found that the susceptibility of MRSA to ceftaroline decreased marginally from 99.4% in 2010 to 98.6 % in 2016 [18].

This study found that the MSSA isolates had much lower ceftaroline MIC's when compared to MRSA isolates. Majority (90.9%) of MSSA isolates had an MIC ≤ 0.25 $\mu\text{g/mL}$. This correlates with the findings from a multi-centric study from India [14]. A multi-centric study from Latin American countries, as part of AWARE surveillance program, also reported a similar finding with 98.3% of MSSA isolates having a ceftaroline MIC of ≤ 0.25 $\mu\text{g/mL}$ [19].

All the MRSA isolates in this study were found to be susceptible to ceftaroline with 0.5 $\mu\text{g/mL}$ being the highest ceftaroline MIC detected. This is in contrast to other studies from India which have reported a higher ceftaroline MIC. Further details are shown in [Table/Fig-6] [14,15].

Authors	n	No. of strains of MRSA (%) inhibited at ceftaroline MIC ($\mu\text{g/mL}$) of:		
		≤ 1	2-4	8
Bakthavatchalam YD et al., [14]	n=86	73 (84.88)	13 (15.12)	Nil
Gaikwad V et al., [15]	n=30	28 (93.33)	2 (6.67)	Nil
Present study	n=28	28 (100)	Nil	Nil

[Table/Fig-6]: Comparison of ceftaroline MIC of MRSA strains in India [14,15].
MIC: Minimum inhibitory concentration; MRSA: Methicillin resistant *staphylococcus aureus*

One such study showed that 6% and 2% of tested *S. aureus* had a ceftaroline MIC of 2 $\mu\text{g/mL}$ and 4 $\mu\text{g/mL}$, respectively [14], which according to the old Clinical and Laboratory Standards Institute (CLSI) guidelines were interpreted as intermediate and resistant respectively. If the current CLSI guidelines are applied, these isolates will be classified as SDD and not as resistant [14,16]. The authors of another study from Maharashtra, India reported that 93.33% MRSA isolates were susceptible to ceftaroline with the MIC being 0.75 $\mu\text{g/mL}$ and concluded that it can be considered an effective alternative treatment, while vancomycin and linezolid can be kept as reserve drug [15]. A multicentric study across seven provinces in Turkey found that 94.3% of tested MRSA isolates were inhibited by ceftaroline (MIC ≤ 1 $\mu\text{g/mL}$) [20]. In a study conducted in the US hospitals from 2008-2011, the authors found that all daptomycin non-susceptible *Staphylococci* isolates, 85.7% and 91.9% of linezolid-resistant *S.aureus* isolates and *S.aureus* isolates with a vancomycin MIC of ≥ 2 $\mu\text{g/mL}$ respectively were susceptible to ceftaroline. The authors concluded that ceftaroline may be considered as a valuable treatment option for infections caused by multidrug resistant *S. aureus* [21].

The recommended dosage of ceftaroline is 600 mg administered every 12 hours by intravenous (IV) infusion over 60 minutes in patient's ≥ 18 years of age [8]. Apart from the clinical efficacy, it is important to consider the adverse effects as well while prescribing any drug. The most common adverse effects with ceftaroline includes nausea, vomiting and diarrhoea. The incidence of which is 3-5% which was comparable with vancomycin/aztreonam with a dosing regimen of vancomycin 1g every 12 hours plus aztreonam 1g every eighth hourly [9]. Clinical trials have also shown that ceftaroline is as effective as ceftriaxone, and combination of vancomycin/

aztreonam for the treatment of community-acquired pneumonia and complicated SSI, respectively [22].

Limitation(s)

This study has evaluated a small number of isolates, as it was an ICMR-STS project with a limited study duration of two months. Prospective studies with larger sample size are warranted to support or verify the findings. Secondly, the present study has not evaluated resistance of MRSA isolates to other antibiotics like vancomycin, linezolid, teicoplanin which may be considered as a limitation of this study.

CONCLUSION(S)

Taking into consideration, the high susceptibility rates and comparable or better tolerance of patients to ceftaroline, when compared to vancomycin, for the treatment of infections caused by MRSA isolates, ceftaroline can be considered as an effective alternative for treatment of infections caused by MRSA. But like most drugs, ceftaroline might become ineffective, if misused. With the increasing resistance to antibiotics and very few newer antibiotics in the pipeline, it is high time to stop misusing the antibiotics in order to help in combating the development of further resistance and prevent going back to the pre-antibiotic era.

REFERENCES

- Indian Network for Surveillance of Antimicrobial Resistance (INSAR) group, India. Methicillin Resistant *Staphylococcus aureus* (MRSA) in India: Prevalence and susceptibility pattern. Indian J Med Res. 2013;137:363-69.
- Al-Hamad AM, Alfaraj AA, Altowailib JA, Al-Shamlan SM, Leskafi H, Alsubeikhy FA, et al. Incidence and antibiotic susceptibility of MRSA infections in a Saudi Arabian Hospital: A 10-year surveillance study. J Infect Dev Ctries. 2018;12(6):454-67.
- Pletz MW, Burkhardt O, Welte T. Nosocomial methicillin -resistant *Staphylococcus aureus* (MRSA) pneumonia: linezolid or vancomycin? - Comparison of pharmacological and clinical efficacy. Eur J Med Res. 2010;15:507-13.
- Cosimi RA, Beik N, Kubiak DW, Johnson JA. Ceftaroline for severe Methicillin -Resistant *Staphylococcus aureus* infections: A systematic review. Open forum infectious diseases. 2017; 4(2): ofx084. doi: 10.1093/ofid/ofx084.
- Jacob JT, Diaz Granados CA. High vancomycin minimum inhibitory concentration and clinical outcomes in adults with methicillin-resistant *Staphylococcus aureus* infections: A meta-analysis. Int J Infect Dis. 2013;17:e93-100. doi: 10.1016/j.ijid.2012.08.005.
- Micek ST. Alternatives to vancomycin for the treatment of methicillin-resistant *Staphylococcus aureus* Infections. CID. 2007;45:S184-90.
- Slusarczyk R, Bielejewska A, Bociek A, Bociek M. Resistance to ceftaroline-2018 review. European Journal of Biological Research. 2018;8(3):112-20.
- Laudano JB. Ceftaroline fosamil: A new broad-spectrum cephalosporin. J Antimicrob Chemother. 2011;66:iii11-iii18.
- Lan SH, Chang SP, Lai CC, Lu LC, Chao CM. Ceftaroline efficacy and safety in treatment of complicated skin and soft tissue infection: A systemic review and meta-analysis of randomized controlled trials. J Clin Med. 2019;8(6):776. doi: 10.3390/jcm8060776.
- Zasowski EJ, Trinh TD, Claeys KC, Casapao AM, Sabagha N, Lagnf AM, et al. Multicenter observational study of ceftaroline fosamil for methicillin-resistant *Staphylococcus aureus* bloodstream infections. Antimicrob Agents Chemother. 2017;61(2):e02015-16. doi: 10.1128/AAC.02015.
- Kelley WL, Joussein A, Barras C, Lelong E, Renzoni A. Missense mutations in PBP2A Affecting ceftaroline susceptibility detected in epidemic hospital-acquired methicillin-resistant *Staphylococcus aureus* clonotypes ST228 and ST247 in Western Switzerland archived since 1998. Antimicrob Agents Chemother. 2015;59(4):1922-30. doi:10.1128/AAC.04068-14.
- Sader HS, Flamm RK, Jones RN. Antimicrobial activity of ceftaroline and comparator agents tested against bacterial isolates causing skin and soft tissue infections and community-acquired respiratory tract infections isolated from the Asia-Pacific region and South Africa. Diagn Microbiol Infect Dis. 2010;76(1):61-68. doi: 10.1016/j.diagmicrobio.2013.01.005.
- Basireddy S, Singh M, Ali S, Kabra V. In vitro activity of ceftaroline against methicillin-resistant *Staphylococcus aureus* isolates. Indian J Med Microbiol. 2015;33:464-65.
- Bakthavatchalam YD, Prasagam AK, Anandan S, Joshi S, Chaudhuri BN, Chitnis DS, et al. Comparative in-vitro activity of Ceftaroline against *Staphylococcus aureus* isolates from India. J Infect Dev Ctries. 2016;10(3):109-12.
- Gaikwad V, Gohel T, Panickar S, Chincholkar V, Mangalkar S. In vitro activity of ceftaroline: A novel antibiotic against methicillin-resistant *Staphylococcus aureus*. Indian J Pathol Microbiol. 2016;59(4):496-98.
- CLSI. Performance Standards for Antimicrobial susceptibility testing. 29th ed. CLSI supplement M100. Wayne PA: Clinical and laboratory standards institute; 2019.
- Tenorio-Abreu A, Gil Tomás J, Bratos Pérez MÁ, de la Iglesia Salgado A, Borrás

- Máñez M, de Lejarazu O, et al. In vitro activity of ceftaroline against Spanish isolates of *Staphylococcus aureus*: A multicenter study. *Enfermedades Infecc Microbiol Clínica*. 2015;33:101-04.
- [18] Sader HS, Mendes RE, Streit JM, Flamm RK. Antimicrobial susceptibility trends among *Staphylococcus aureus* isolates from U.S. hospitals: Results from 7 years of the ceftaroline (AWARE) surveillance program, 2010 to 2016. *Antimicrob Agents Chemother*. 2017;61:e01043-17. <https://doi.org/10.1128/AAC.01043-17>.
- [19] Biedenbach DJ, Hoban DJ, Reiszner E, Lahiri SD, Alm RA, Sahm DF, et al. In vitro activity of ceftaroline against *Staphylococcus aureus* isolates collected in 2012 from Latin American countries as part of the AWARE surveillance program. *Antimicrob Agents Chemother*. 2015; 59(12):7873-77. doi: 10.1128/AAC.01833-15
- [20] Mengeloglu FZ, Taş T, Koçoglu E, Copur Çiçek A, Yanık K, Güneş H, et al. In vitro activity of ceftaroline to MRSA isolates: A multicenter study. *Mikrobiyol Bul*. 2013;47:677-83.
- [21] Sader HS, Flamm RK, Jones RN. Antimicrobial activity of ceftaroline tested against *Staphylococci* with reduced susceptibility to linezolid, daptomycin, or vancomycin from U.S. hospitals, 2008 to 2011. *Antimicrob Agents Chemother*. 2013;57:3178-81. doi:10.1128/AAC.00484-13.
- [22] Henry P, Mei HC, Horatio BF. Ceftaroline fosamil: A cephalosporin with activity against methicillin-resistant *Staphylococcus aureus*. *Clinical Therapeutics*. 2012;34(4):743-65.

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